

THE INTERNATIONAL JOURNAL OF HUMANITIES & SOCIAL STUDIES

Impact on Rheological Properties of Fermented Milk Products Due to Lactic Acid Bacteria

Kele VD.

College of Dairy Technology, Udgir, District Latur, Maharashtra, India

Behare PV.

College of Dairy Technology, Udgir, District Latur, Maharashtra, India

Bajad DN.

College of Dairy Technology, Udgir, District Latur, Maharashtra, India

Abstract:

EPS producing lactic cultures have tremendous potential as functional starters, which can be better substituted to many commercial additives in use. They have significant effect on rheological and organo-leptic properties of a food. Incorporation of these cultures in to food products not only satisfy consumer demands of natural, healthy and low calorie food product but also provide food with as few food additives as possible. Use of EPS producing cultures reduce the amount of total solids required to manufacture dahi, lassi and yoghurt and thus making the process cost effective. Fermented milks, especially made by traditional method in rural areas, are the potential reservoir for isolation of EPS producing cultures for Consumers' demand for natural, healthy and low-calorie food has increased interest in the dairy industries for development and manufacture of low-fat/fat-free fermented milk products.

Keywords: EPS, commercial additives, dairy, fermented milk product, consumer

1. Introduction

Consumers' demand for natural, healthy and low-calorie food has increased interest in the dairy industries for development and manufacture of low-fat/fat-free fermented milk products. But fat removal has several undesirable effects on physical properties of fermented milks such as inferior flavour, texture and rheo-logical properties that consequently hamper their acceptability. Several attempts were made including increase in milk solids and addition of stabilizers to tackle such problems (Rohm and Schmid 1993). However, these approaches did not address an increasing consumer demand for products with natural, low-cost and with as few food additives as possible. Furthermore, additives particularly, stabilizers are strictly prohibited in some fermented milk products like dahi (similar to yoghurt) in India and yoghurt in Norway by the stringent regulations. In this con- text, there is no alternative to use EPS producing lactic cultures, which offer a natural and usually acceptable way for making low-fat/fat-free fermented milks Lactic acid bacteria (LAB) are widely used in the dairy and food industry since time immemorial. Apart from production of lactic acid, flavouring compounds and bacteriocin like substances, several strains of LAB secrete extracellular polysaccharide in favourable environment such as milk (Cerning and Marshall 1999). The term exopolysaccharide (EPS) is used to describe extracellular polysaccharide either attached as capsule with bacterial cell wall or liberated into the medium as ropy polysaccharide (Sutherland 1972). The EPS play an important role in the improvement of physical properties of fermented milks, which act like a food stabilizer, viscosifier, emulsifier or gelling agent providing a product with natural thickness (Ruas-Madiedo and Reyes-Gavilan 2005). Some of the examples of LAB EPS are dextran, mutan and fructan produced by *Leuconostocmesenteroides*, *Streptococcus mutans* and *Strep. salivariussubsp. thermophilus*, respectively (Montiville et al 1978, Cerning 1990). The Gram negative bacteria, *Xanthomonascampetris*, *Acetobacterxylinum* and *Sphingomonaspaucimovilis* also produce EPS xanthan, acetan and gellan, respectively that are commercially available as food additives (Harvey and McNeil 1998). However, EPS extracted from Gram negative bacteria, although produced in larger quantities than EPS produced by food grade LAB, may not be preferred, as former is derived from non-food grade non-GRAS (Generally Recognized As Safe) status organism and had high cost involved in their recovery. Moreover, addition of purified EPS into the food product may not have similar effects as EPS produced *in situ* by LAB during milk fermentation (Doleyreset al 2005). The properties of EPS in purified form differ considerably from the properties of EPS produced *in situ* (Dubocand Mollet 2001), latter being a more desirable approach. The *in situ* EPS production may play useful role in the manufacture of a variety of cultured dairy products such as yoghurt, drinking yoghurt, cheese, cultured cream and milk-based dessert (Crescenzi 1995, Bouzar et al 1997,).

A large variety of LAB, with some strains of Bifidobacteria are reported to produce EPS (Zotta et al 2008). Most of them belong to the genera of *Lactococcus*, *Streptococcus*, *Lactobacillus*, *Leuconos-toc* and *Pediococcus*. The use of polysaccharide producing lactic culture strains in the fermented milk manufacture is not new. The EPS producing LAB have been traditionally used in the Scandina-

vian fermented milk products to impart desirable texture and rheological properties (Macura and Townsley 1984). The products made with ropy strains have smooth body, high viscosity and less syneresis than the products made with non-ropy strains (Wacher-Rodarte et al 1993). The EPS producing lactic bacteria are isolated from dairy and non-dairy environment using different media supplemented with one or more type of sugars. The media used for isolation of EPS producing cultures are: liquid EPS selection medium (ESM) containing (g/l) 90 skim milk, 3.5 yeast extract, 3.5 peptone and 10 glucose (Van den Berg et al 1993), milk indicator agar and M17 lactose agar (Terzaghi and Sandine 1975), MRS with high concentration of sugars (100/g) (Van Geel-Schutten et al 1998) and milk agar (Mozzi et al 2001).

Milk fat contributes to the body, texture and flavour development of dairy products. Fat reduction to satisfy consumers demand leads to textural and functional defects in low-fat yoghurt, cheeses, *dahi*, and kefir (Mistry 2001, Guven et al 2005). In low-fat yoghurt and *dahi*, lack of flavour, weak body and poor texture are the major problems. Mechanical breaking particularly in stirred yoghurt, strongly affects the rheology of the coagulum and favour syneresis since the network formed by the gel is broken (Duboc and Mollet 2001). Low-fat cheeses have poor moisture retention ability, which otherwise moisture present in the cheese partially overcomes the problem of firm, rubbery body and texture created by high casein content (Mistry 2001). Generally for cheese manufacture, the casein to fat ratio is 0.68-0.70, which gives desirable textural attributes. Due to fat reduction, the casein to fat ratio is disturbed and casein content is more in the resultant cheese consequently generating some textural defects. Low-fat Mozzarella cheeses have less tendency to melt and inferior baking characteristics (Fife et al 1996). These defects can be reduced by the use of additives to some extent but may not find wide acceptance to produce wholesome product. The consistent manufacture of good quality products that have good texture, mouthfeel and stability is important to the dairy industry. The manufacturers have used texture promoting or ropy cultures for many years particularly where addition of stabilizer is prohibited (Marshall and Rawson 1999). These cultures may impart higher intensity to flavour to the yoghurt due to the carbohydrate masking the flavour, mouth feel and other attributes (Tamime and Robinson 1999). To reduce the amount of added milk solids, improve yoghurt viscosity, to enhance texture and mouthfeel and to avoid syneresis during fermentation or upon storage of the fermented milk products, EPS producing functional starters are interesting (De Vuyst et al 2003).

The apparent viscosity of skim milk gel made by both ropy cultures increased as compared to that made by non-ropy cultures (Tamime and Robinson 1999). However, combining two ropy cultures for yoghurt manufacture may not have always, additive effect (Faber et al 1998). Marshall and Rawson (1999) found that mixing a non-ropy strain of *Strep. thermophilus* with a ropy strain of *Lact. delbrueckii* subsp. *bulgaricus* had a greater effect on viscosity of stirred yoghurt than combining 2 ropy strains. The authors suggested the interaction and cooperative growth that occur in mixed cultures, which also appear to influence EPS production in yoghurt. The presence of EPS in stirred yoghurt makes the product less susceptible to mechanical damage from pumping, blending and filling machines (Robinson 1981). Although, mechanical processing steps do increase the syneresis of the final product, use of EPS cultures can control this defect (Duboc and Mollet 2001). Hassan et al (1996a, b) examined the rheological and textural properties of yoghurt made with strains differentiated as encapsulated non-ropy (in which an EPS capsule is formed), encapsulated ropy (secretes extracellular slime) and non-encapsulated non-ropy. Yoghurt made with ropy cultures exhibited increased viscosity and shear stress values; however, differences attributed to the type of polysaccharide secretion (capsule or slime) were apparent. The presence of bacterial capsule may enhance some rheological properties such as viscosity, but may weaken the gel structure. This has caused lower shear stress value compared to slime producers, which produce a more stretchable gel structure (Hassan et al 1996a). The types of EPS produced by the yoghurt bacteria have effects on the texture and syneresis. Yoghurt made with encapsulated non-ropy cultures had the lowest firmness and curd tension, but exhibited less syneresis than unencapsulated cultures. The lower firmness in the yoghurt made by slime producing cultures might have been due to the polysaccharide interfering with the casein structure (Hassan et al 1996b). Several microstructural studies have indicated that it is not the amount of EPS, which is important to the rheological properties of fermented milk but the type of EPS and therefore the interaction of the polymer with the bacterial cell and the milk protein is important during the fermentation (Marshall and Rawson 1999). Folkenberg et al (2005) observed 2 types of microstructure in yoghurt made with different EPS producing cultures, one in which EPS is associated with the protein network and another where EPS appeared to be incompatible with the protein. Yoghurt in which the EPS were associated with protein had ropiness, low serum separation and appeared more resistant to stirring than EPS appeared incompatible with protein.

Incorporation of EPS strains in the starter culture retains significant amount of moisture in a variety of low-fat cheeses that positively influence their functionality (Awad et al 2005, Zisu and Shah 2005). Perry et al (1997) investigated the influence of an EPS-producing starter pair *Strep. thermophilus* MR-1C and *Lact. delbrueckii* subsp. *bulgaricus* MR-1R on the moisture and melt properties of low-fat Mozzarella cheese. Cheese manufactured with MR-1C and MR-1R contained significantly ($p < 0.05$) more moisture and had better melting properties than cheese made with an EPS negative commercial starter pair (*Strep. thermophilus* TA061 and *Lact. helveticus* LH 100). The water binding properties of *Strep. thermophilus* MR-1C was ascribed to its large capsule. Cheddar cheese made with EPS-producing *Lact. lactis* retained more moisture (45.8-47.2%) than control made with no EPS producing strains (44.9-45.8%). The texture profile analysis revealed that cohesiveness, chewiness and tortion shear strain values of the *Lact. lactis* cheeses were lower than those of control and the meltability and springiness values were higher because EPS contained cheese had a more open structure, which is more conducive to mastication and melting (Tunick et al 2003). EPS producing cultures improved textural, melting and sensory characteristics of reduced fat cheddar cheese (Awad et al 2005). A ropy *L. lactis* subsp. *cremoris* strain produced reduced-fat cheddar cheese with a moisture in the non-fat substances level and textural and melting properties similar to those of the full-fat type due to the ability of EPS to bind significant amounts of free water. Nauth and Hayashi (2004) patented the process of manufacture of fat-free cream cheese, added with ropy culture. The fat-free cream cheese has comparable firmness, consistency and flavour of a conventional higher fat cream cheese.

Besides yoghurt and cheese, the other fermented milk products in which EPS cultures have been shown to affect physical properties are sour cream, kefir and European cultured dairy products. Use of slime producing *Strep. thermophilus* strains greatly improved

rheological properties of cream turo and number of other Hungarian cultured milk and cultured cream products (Obert 1984). Kefir is traditional self-carbonated slightly alcoholic fermented milk from Eastern Europe (Roginski 1999, Tamime and Robinson 1999). Kefir is prepared by inoculating kefir grains, which consists of homofermentative, heterofermentative LAB, yeasts and acetic acid bacteria. The cells are embedded in kefiran, a slimy polysaccharide, which also found to affect texture of kefir (Micheli et al 1999).

2. Conclusion

Food and Dairy industry is looking for the multifunctional strains of LAB that contribute to the organoleptical, technological, nutritional and health properties of fermented milk products. EPS producing lactic cultures have tremendous potential as functional starters, which can be better substituted to many commercial additives in use. They have significant effect on rheological and organoleptic properties of a food. Incorporation of these cultures in to food products not only satisfy consumer demands of natural, healthy and low calorie food product but also provide food with as few food additives as possible. Use of EPS producing cultures reduce the amount of total solids required to manufacture dahi, lassi and yoghurt and thus making the process cost effective. Fermented milks, especially made by traditional method in rural areas, are the potential reservoir for isolation of EPS producing cultures.

3. References

1. Awad, S, Hassan N, Muthukumarappan K 2005. Application of exopolysaccharide-producing cultures in reduced fat cheddar cheese: Texture and melting properties. *J Dairy Sci* 88:4204-4213
2. Bouzar F, Cerning J, Desmazeaud M 1997. Exopolysaccharide production and texture promoting abilities of mixed-strain starter cultures in yoghurt production. *J Dairy Sci* 80:2310-2317
3. Cerning J 1990. Exocellular polysaccharides produced by lactic acid bacteria. *FEMS Microbiol Rev* 87:113-130
4. Cerning J, Marshall VM 1999. Exopolysaccharides produced by the dairy lactic acid bacteria. *Recent Res Develop Microbiol* 3:195-209
5. Crescenzi V 1995. Microbial polysaccharides of applied interest: On going research activities in Europe. *Biotechnol Progr* 11:251-259
6. De Vuyst L, Zamfir M, Mozzi F, Adriany T, Marshall V, Degeest B, Vaningelgem F 2003. Exopolysaccharide-producing *Streptococcus thermophilus* strains as functional starter cultures in the production of fermented milks. *Int Dairy J* 13:707-717
7. Doleyres Y, Schaub L, Lacroix C 2005. Comparison of the functionality of exopolysaccharides produced in situ or added as bioingredients on yoghurt properties. *J Dairy Sci* 88:4146-4156
8. Duboc P, Mollet B 2001. Applications of exopolysaccharides in dairy industry. *Int Dairy J* 11:759-768
9. Faber EJ, Zoon P, Kamerling JP, Vliegthart JFG 1998. The exopolysaccharides produced by *Streptococcus thermophilus* strains have the same repeating unit but differ in viscosity of their milk cultures. *Carbohydr Res* 310:269-276
10. Fife RL, McMahon DJ, Oberg CJ 1996. Functionality of low fat Mozzarella cheese. *J Dairy Sci* 79:1903-1910
11. Folkenberg DM, Dejmeck P, Skriver A, Ipsen R 2005. Relation between sensory texture properties and exopolysaccharide distribution in set and in stirred yoghurts produced with different starter cultures. *J Texture Stud* 36:174-189
12. Guven M, Yasar K, Karaca OB, Hayaloglu AA 2005. The effect of inulin as a fat replacer on the quality of set-type low-fat yoghurt manufacture. *Int J Dairy Technol* 58:180-184
13. Harvey LM, McNeil B 1998. Thickeners of microbial origin. In: *Microbiology of fermented foods*. Wood J B (ed), Blackie Academic and Professional, Glasgow, UK, p 148-171
14. Hassan AN, Frank JF, Schmidt KA, Shalabi SI 1996a. Textural properties of yoghurt made with encapsulated non-ropy lactic cultures. *J Dairy Sci* 79:2098-2103
15. Hassan AN, Frank JF, Schmidt KA, Shalabi SI 1996b. Rheological properties of yoghurt made with encapsulated non-ropy lactic cultures. *J Dairy Sci* 79:2091-2097
16. Macura D, Townsley PM 1984. Scandinavian ropy milk: Identification and characterization of endogenous ropy lactic streptococci and their extracellular excretion. *J Dairy Sci* 67:735-744
17. Marshall VM, Rawson HL 1999. Effects of exopolysaccharide-producing strains of thermophilic lactic acid bacteria on the texture of stirred yoghurt. *Int J Food Sci Technol* 34:137-143
18. Mistry VV 2001. Low-fat cheese technology. *Int Dairy J* 11:413-422
19. Montville TH, Cooney CL, Sinskey AJ 1978. *Streptococcus mutans* dextranase: A review. *Adv Appl Microbiol* 24:55-84
20. Mozzi F, Rollan G, Savoy de Giori G, Font de Valdez G 2001. Effect of galactose and glucose on the exopolysaccharide production and the activities of biosynthetic enzymes in *Lactobacillus casei* CRL 87. *J Appl Microbiol* 91:160-167
21. Nauth RK, Hayashi D 2004. Methods for manufacture of fat-free cream cheese. US Patent US 6,689,402
22. Nauth RK, Hayashi D 2004. Methods for manufacture of fat-free cream cheese. US Patent US 6,689,402
23. Obert G 1984. Manufacturing technology of cream turo with extended shelf-life, and isolation of a special strain of lactic acid bacteria which improves product consistency. *Tejipar* 33:47-48
24. Perry DB, McMahon DJ, Oberg CJ 1997. Effect of exopolysaccharide producing cultures on moisture retention in low-fat Mozzarella cheese. *J Dairy Sci* 80:799-805
25. Robinson DK 1981. Yoghurt manufacture-some consideration of quality. *Dairy Ind Int* 46:31-35
26. Roginski H 1999. Fermented milks: Product from Northern Europe. In: *Encyclopedia of food microbiology*. Robinson RK (ed), Academic Press, San Diego. p 791-798

27. Rohm H, Schmid W 1993. Influence of dry matter fortification on flow properties of yoghurt. 1. Evaluation of flow curves. *Milchwissenschaft* 48:556-560
28. Ruas-Madiedo P, de los Reyes-Gavilan CG 2005. Methods for the screening, isolation, and characterization of exopolysaccharides produced by lactic acid bacteria. *J Dairy Sci* 88:843-856
29. Sutherland IW 1972. Bacterial exopolysaccharides. *AdvMicrobPhysiol* 8:143-212
30. Tamime AY, Robinson RK 1999. *Yoghurt: Science and technology*. CRC Press, Cambridge
31. Terzaghi BE, Sandine WE 1975. Improved medium for lactic Streptococci and their bacteriophages. *Appl Environ Microbiol* 29:807-813
32. Tunick MH, Van Hekken DL, Malin EL, Guinee TP, Beresford TP, Broadbent J, McMahon DJ 2003 Rheology of low-fat cheddar cheeses made with exopolysaccharide-producing cultures *Am ChemSoc Paper Nr AGF 0063* (abstract).
33. Van den Berg DJC, Robijn GW, Janssen AC, Giuseppin MLF, Vreeker R, Kamerling JP, Vliegthart JFG, Ledebor AM, VerripsCT 1995. Production of a novel extracellular polysaccharide by *Lactobacillus sake* 0-1 and characterization of the polysaccharide. *Appl Environ Microbiol* 61:2840-2844
34. Van Geel-Schuten GH, Flesch F, Brink Bten, Smith MR, Dijkhuizen L 1998. Screening and characterization of *Lactobacillus* strains producing large amounts of exopolysaccharides. *ApplMicrobiolBiotechnol* 50:697-703
35. Wachter-Rodarte CM, Galvan MV, Farres A, Gallardo F, Marshall VME, Garcia-GaribayM 1993. Yoghurt production from reconstituted skim milk powders using different polymer and non-polymer forming starter cultures. *J Dairy Res* 60:247-254
36. Zisu B, Shah NP 2005. Low-fat Mozzarella as influenced by microbial exopoly saccharides, preacidification and whey protein concentrate. *J Dairy Sci* 88:1973-1985
37. Zotta T, Piraino P, Parente E, Salzano G, Ricciardi A 2008. Characterization of lactic acid bacteria isolated from sourdoughs for Cornetto, a traditional bread produced in Basilicata (Southern Italy). *World J MicrobiolBiotechnol* (accepted on 17 January 2008, Online available www.springerlink.com/inline/ex/e66367118k10km17.pdf)